Quick Fly Genomic DNA prep

1) Collect 30 anesthetized flies in Eppendorf tube and freeze at -80°.
2) Grind flies in 200µL Buffer A with disposable tissue grinder (Kontes).
3) Add an additional 200µL Buffer A and continue grinding until only cuticles remain.
4) Incubate at 65° for 30 minutes.
5) Add 800µL LiCl/KAc Solution and incubate on ice for at least 10 minutes.
6) Spin for 15 minutes at RT.
7) Transfer 1mL of the supernatant into a new tube, avoiding floating crud.
8) [If crud transfers, respin.]
9) Add 600µL isopropanol (0.6 volume of S/N), mix, spin 15 minutes at RT.
   As extraction does not remove all the proteins, do not incubate on ice and spin immediately after addition of isopropanol.
10) Aspirate away supernatant, pulse, aspirate and wash with 70% ethanol, dry.
11) Resuspend in 150µL 10mM Tris pH ~ 7 (e.g. .Buffer EB from QIAGEN plasmid kits)
12) Store at -20°.

Buffer A

100 mM Tris-HCl, pH 7.5
100 mM EDTA
100 mM NaCl
1% SDS

LiCl/KAc Solution

1 part 5 M KAc stock
2.5 parts 6 M LiCl stock